

First published online as a Review in Advance on April 17, 2007

Effects of Brain Evolution on Human Nutrition and Metabolism

William R. Leonard,¹ J. Josh Snodgrass,² and Marcia L. Robertson¹

¹Department of Anthropology, Northwestern University, Evanston, Illinois 60208, ²Department of Anthropology, University of Oregon, Eugene, Oregon 97403; email: w-leonard1@northwestern.edu

Annu. Rev. Nutr. 2007. 27:311–27

The Annual Review of Nutrition is online at http://nutr.annualreviews.org

This article's doi: 10.1146/annurev.nutr.27.061406.093659

Copyright © 2007 by Annual Reviews All rights reserved

0199-9885/07/0821-0311\$20.00

Key Words

encephalization, hominin, diet quality, body composition, *Homo erectus*

Abstract

The evolution of large human brain size has had important implications for the nutritional biology of our species. Large brains are energetically expensive, and humans expend a larger proportion of their energy budget on brain metabolism than other primates. The high costs of large human brains are supported, in part, by our energy- and nutrient-rich diets. Among primates, relative brain size is positively correlated with dietary quality, and humans fall at the positive end of this relationship. Consistent with an adaptation to a high-quality diet, humans have relatively small gastrointestinal tracts. In addition, humans are relatively "undermuscled" and "over fat" compared with other primates, features that help to offset the high energy demands of our brains. Paleontological evidence indicates that rapid brain evolution occurred with the emergence of *Homo erectus* 1.8 million years ago and was associated with important changes in diet, body size, and foraging behavior.

Contents

INTRODUCTION	312
COMPARATIVE PERSPECTIVES	
ON BRAIN SIZE, BODY SIZE,	
DIET, AND METABOLIC	
RATE	312
EVOLUTIONARY CHANGES IN	
BRAIN SIZE AND DIET	315
Trends in the Hominin Brain Size,	
Body Size, and Tooth Size	315
Dietary Changes Associated with	
Brain Evolution in Early	
Homo	317
BRAIN EVOLUTION AND	
HUMAN BODY	
COMPOSITION	319
CONCLUSIONS	321

INTRODUCTION

Over the past 20 years, the evolution of human nutritional requirements has received ever greater attention among both anthropologists and nutritional scientists (3, 21, 28, 29, 35, 48, 49, 86). Increasingly, we have come to understand that many of the key features that distinguish humans from other primates (e.g., our bipedal form of locomotion and large brain sizes) have important implications for our distinctive nutritional needs (3, 47, 50). The most important of these is our high levels of encephalization (large brain:body mass). The energy demands (kcal/g/min) of brain and other neural tissues are extremely highapproximately 16 times that of skeletal muscle (37, 43). Consequently, the evolution of large brain size in the human lineage came at a very high metabolic cost.

Despite the fact that humans have much larger brains per body weight in comparison with other primates or terrestrial mammals, the resting energy demands for the human body are no more than for any other mammal of the same size (48, 49). The consequence of this paradox is that humans allocate a much larger share of their daily energy budget to "feed their brains." Brain metabolism accounts for $\sim 20\%$ to 25% of resting metabolic rate (RMR) in an adult human body. This is far more than the 8% to 10% observed in other primate species and still more than the 3% to 5% allocated to the brain by other (nonprimate) mammals (49).

The disproportionately large allocation of our energy budget to brain metabolism has important implications for our dietary needs. This review draws on both analyses of living primate species and the human fossil record to examine the avenues through which humans have adapted to the metabolic demands of greater encephalization. We begin by considering the energy demands associated with large brain size in modern humans relative to other primates and nonprimate mammals. Next we examine comparative dietary data for modern human groups and other primate species to evaluate the influence that variation in relative brain size has on dietary patterns among modern primates. We then turn to an examination of the human fossil record to examine when and under what conditions in our evolutionary past key changes in brain size and diet likely took place. Finally, we explore how the evolution of large human brains was likely accommodated by differential changes in the relative sizes of other organs [e.g., muscle, fat, and gastrointestinal (GI) tract]. The high metabolic costs of our large brains appear to play a strong hand in shaping distinctive aspects of human growth and development.

COMPARATIVE PERSPECTIVES ON BRAIN SIZE, BODY SIZE, DIET, AND METABOLIC RATE

Table 1 presents comparative data on RMR, brain size, body size, and diet for living humans and nonhuman primates (from 51). Primates, as a group, are similar to other mammals in having RMRs that scale to approximately three-fourths power of body mass (see 44). **Figure 1** (see color insert) presents the relationship RMR (kcal/day) and body

Encephalization: brain size in relation to body size. In general, primates are more encephalized than other mammals

RMR: resting metabolic rate

GI: gastrointestinal

Species RMR (kcal/d) Body mass (kg) Brain mass (g) Body mass (kg) Alouatta palliata 231.9 4.670 51 6.400 Atus trivirgatus 52.4 1.020 16 0.850 Arctocebus calabarensis 15.2 0.206 7.2 0.323 Callitbrix geoffroyi 27.0 0.225 7.6 0.280 Callitbrix jacebus 22.8 0.356 7.6 0.280 Celuella pygmaea 10.1 0.105 4.5 0.140 Cercopithecus mitis 407.7 8.500 76 6.500 Cercocebus torquatus 196.2 4.000 104 7.900 Cheirogaleus medius 22.7 0.300 3.1 0.177 Colobus guereza 357.9 10.450 73 7.000 Erytbrocebus patas 186.9 3.000 118 8.000 Eulerus dugamulus 25.1 0.260 7.2 0.274 Galago senegalensis 18.1 0.215 4.8 0.186 <th>DQ 136 177.5 327.5 235 249.5 249.5 249.5 234 126</th>	DQ 136 177.5 327.5 235 249.5 249.5 249.5 234 126
Alonatta palliata 231.9 4.670 51 6.400 Aotus trivirgatus 52.4 1.020 16 0.850 Arctocebus calabarensis 15.2 0.206 7.2 0.323 Callitbrix gachus 22.8 0.356 7.6 0.280 Callitbrix jacebus 22.8 0.356 7.6 0.280 Cellula pygmaea 10.1 0.105 4.5 0.140 Cercopitbecus mitis 407.7 8.500 76 6.500 Cercocbus torquatus 196.2 4.000 104 7.900 Cheirogaleus medius 22.7 0.300 3.1 0.177 Colobus guereza 357.9 10.450 73 7.000 Erytbroebus patas 186.9 3.000 118 8.000 Eulemur fulvus 42.0 2.397 25.2 2.397 Euoticus elegantulus 25.1 0.260 7.2 0.274 Galago senegalensis 18.1 0.215 4.8 0.186 Galago	136 177.5 327.5 235 249.5 249.5 201.5 234 126
Actus trivirgatus 52.4 1.020 16 0.850 Arctocebus calabarensis 15.2 0.206 7.2 0.323 Callitbrix geoffroyi 27.0 0.225 7.6 0.280 Callitbrix jacebus 22.8 0.356 7.6 0.280 Celulta pygmaca 10.1 0.105 4.5 0.140 Cercopitbecus mitis 407.7 8.500 76 6.500 Cercopitbecus mitis 407.7 8.500 76 6.500 Cercocebus torquatus 196.2 4.000 104 7.900 Cherosgaleus medius 22.7 0.300 3.1 0.177 Colobus guereza 357.9 10.450 73 7.000 Erythrocebus patas 186.9 3.000 118 8.000 Eulemur fulcus 25.1 0.260 7.2 0.274 Galago moboli 13.9 0.155	177.5 327.5 235 235 249.5 201.5 234 126
Arctocebus calabarensis 15.2 0.206 7.2 0.323 Callitbrix geoffroyi 27.0 0.225 7.6 0.280 Callitbrix jaccbus 22.8 0.356 7.6 0.280 Callitbrix jaccbus 22.8 0.356 7.6 0.280 Cebuella pygmaea 10.1 0.105 4.5 0.140 Cercopithecus mitis 407.7 8.500 76 6.500 Cercocebus torquatus 196.2 4.000 104 7.900 Cbeirogaleus medius 22.7 0.300 3.1 0.177 Colobus guereza 357.9 10.450 73 7.000 Erytbroecbus patas 186.9 3.000 118 8.000 Eulemur fulcus 42.0 2.397 25.2 2.397 Euoticus elegantulus 25.1 0.260 7.2 0.274 Galago senegalensis 18.1 0.215 4.8 0.186 Galago senegalensis 1400.0 53.500 1295 53.500	327.5 235 235 249.5 201.5 234 126
Callitbrix geoffroyi 27.0 0.225 7.6 0.280 Callitbrix jacchus 22.8 0.356 7.6 0.280 Cellitbrix jacchus 22.8 0.356 7.6 0.280 Cebuella pygmaea 10.1 0.105 4.5 0.140 Cercopithecus mitis 407.7 8.500 76 6.500 Cercocebus torquatus 196.2 4.000 104 7.900 Cheirogaleus medius 22.7 0.300 3.1 0.177 Colobus guereza 357.9 10.450 73 7.000 Erytbroecbus patas 186.9 3.000 118 8.000 Eulemur fulvus 42.0 2.397 25.2 2.397 Euoticus elegantulus 25.1 0.260 7.2 0.274 Galago moboli 13.9 0.155	235 235 249.5 201.5 234 126
Callitbrix jacebus 22.8 0.356 7.6 0.280 Cebuella pygmaea 10.1 0.105 4.5 0.140 Cercopitbecus mitis 407.7 8.500 76 6.500 Cercopitbecus mitis 196.2 4.000 104 7.900 Cheirogaleus medius 22.7 0.300 3.1 0.177 Colobus guereza 357.9 10.450 73 7.000 Erytbrocebus patas 186.9 3.000 118 8.000 Eulemur fulvus 42.0 2.397 25.2 2.397 Euoticus elegantulus 25.1 0.260 7.2 0.274 Galago moboli 13.9 0.155	235 249.5 201.5 234 126
Cebuella pygmaea 10.1 0.105 4.5 0.140 Cercopithecus mitis 407.7 8.500 76 6.500 Cercopithecus mitis 196.2 4.000 104 7.900 Cheirogaleus medius 22.7 0.300 3.1 0.177 Colobus guereza 357.9 10.450 73 7.000 Erythrocebus patas 186.9 3.000 118 8.000 Eulemur fulvus 42.0 2.397 25.2 2.397 Euoticus elegantulus 25.1 0.260 7.2 0.274 Galago moboli 13.9 0.155	249.5 201.5 234 126
Cercopithecus mitis 407.7 8.500 76 6.500 Cercocebus torquatus 196.2 4.000 104 7.900 Cheirogaleus medius 22.7 0.300 3.1 0.177 Colobus guereza 357.9 10.450 73 7.000 Erytbrocebus patas 186.9 3.000 118 8.000 Eulemur fulvus 42.0 2.397 25.2 2.397 Euoticus elegantulus 25.1 0.260 7.2 0.274 Galago moboli 13.9 0.155	201.5 234 126
Cercocebus torquatus 196.2 4.000 104 7.900 Cheirogaleus medius 22.7 0.300 3.1 0.177 Colobus guereza 357.9 10.450 73 7.000 Erytbrocebus patas 186.9 3.000 118 8.000 Eulemur fulvus 42.0 2.397 25.2 2.397 Euoticus elegantulus 25.1 0.260 7.2 0.274 Galago moboli 13.9 0.155	234 126
Cheirogaleus medius 22.7 0.300 3.1 0.177 Colobus guereza 357.9 10.450 73 7.000 Erytbrocebus patas 186.9 3.000 118 8.000 Ealemur fulvus 42.0 2.397 25.2 2.397 Euoticus elegantulus 25.1 0.260 7.2 0.274 Galago moboli 13.9 0.155 Galago senegalensis 18.1 0.215 4.8 0.186 Galago senegalensis 18.1 0.215 4.8 0.081 Homo sapiens 1400.0 53.500 1295 53.500 Hylobates lar 123.4 1.900 102 6.000 Lemur catta 45.1 2.678 25.6 2.678 Leotopitbecus rosalia 51.1 0.718 Lepilemur ruficaudatus 27.6 0.682 7.6 0.682 Loris tardigradus 14.8 0.284 6.6 0.322 Macaca fuscata	126
Colobus guereza 357.9 10.450 73 7.000 Erytbrocebus patas 186.9 3.000 118 8.000 Eulemur fulvus 42.0 2.397 25.2 2.397 Euoticus elegantulus 25.1 0.260 7.2 0.274 Galago moboli 13.9 0.155	126
Erytbrocebus patas 186.9 3.000 118 8.000 Eulemur fulvus 42.0 2.397 25.2 2.397 Euoticus elegantulus 25.1 0.260 7.2 0.274 Galago moboli 13.9 0.155	
Eulemur fulvus 42.0 2.397 25.2 2.397 Euoticus elegantulus 25.1 0.260 7.2 0.274 Galago moboli 13.9 0.155 Galago senegalensis 18.1 0.215 4.8 0.186 Galago senegalensis 18.1 0.215 4.8 0.081 Homo sapiens 1400.0 53.500 1295 53.500 Hylobates lar 123.4 1.900 102 6.000 Lemur catta 45.1 2.678 25.6 2.678 Leontopitbecus rosalia 51.1 0.718 Macaca fascicularis 400.9 7.100 74 5.500 Macaca fuscata 485.4 9.580 84 5.900 Macaca fuscata 445.4 9.580 84 5.900 Macaca fuscata 485.4 9.580 84 5.900 Macaca fuscata 445.4 9.580 110 8.000 Microcebus murinus 4.9	
Euoticus elegantulus 25.1 0.260 7.2 0.274 Galago moboli 13.9 0.155	129
Galago moboli 13.9 0.155 Galago senegalensis 18.1 0.215 4.8 0.186 Galagoides demidoff 6.3 0.058 3.4 0.081 Homo sapiens 1400.0 53.500 1295 53.500 Hylobates lar 123.4 1.900 102 6.000 Lemur catta 45.1 2.678 25.6 2.678 Leontopithecus rosalia 51.1 0.718 1 1 Lepilemur ruficaudatus 27.6 0.682 7.6 0.682 Loris tardigradus 14.8 0.284 6.6 0.322 Macaca fuscata 485.4 9.580 84 5.900 Macaca fuscata 485.4 9.580 84 5.900 Macaca fuscata 485.4 9.580 110 8.000 Microcebus murinus 4.9 0.054 1.8 0.054 Nycticebus coucang 32.4 1.380 12.5 0.800 Otolemur crassicaudatus 47.6 0.950 10.3 0.850	230
Galago senegalensis 18.1 0.215 4.8 0.186 Galagoides demidoff 6.3 0.058 3.4 0.081 Homo sapiens 1400.0 53.500 1295 53.500 Hylobates lar 123.4 1.900 102 6.000 Lemur catta 45.1 2.678 25.6 2.678 Leontopithecus rosalia 51.1 0.718 102 100 Lepilemur ruficaudatus 27.6 0.682 7.6 0.682 Loris tardigradus 14.8 0.284 6.6 0.322 Macaca fascicularis 400.9 7.100 74 5.500 Macaca fuscata 485.4 9.580 84 5.900 Macaca fuscata 485.4 9.580 110 8.000 Microcebus murinus 4.9 0.054 1.8 0.054 Nycticebus coucang 32.4 1.380 12.5 0.800 Otolemur crassicaudatus 47.6 0.950 10.3 0.850	
Galagoides demidoff 6.3 0.058 3.4 0.081 Homo sapiens 1400.0 53.500 1295 53.500 Hylobates lar 123.4 1.900 102 6.000 Lemur catta 45.1 2.678 25.6 2.678 Leontopithecus rosalia 51.1 0.718 1295 1295 Lepilemur ruficaudatus 27.6 0.682 7.6 0.682 Loris tardigradus 14.8 0.284 6.6 0.322 Macaca fascicularis 400.9 7.100 74 5.500 Macaca fuscata 485.4 9.580 84 5.900 Macaca mulatta 231.9 5.380 110 8.000 Microcebus murinus 4.9 0.054 1.8 0.054 Nycticebus coucang 32.4 1.380 12.5 0.800 Otolemur crassicaudatus 47.6 0.950 10.3 0.850	278
Homo sapiens 1400.0 53.500 1295 53.500 Hylobates lar 123.4 1.900 102 6.000 Lemur catta 45.1 2.678 25.6 2.678 Leontopithecus rosalia 51.1 0.718 1295 1295 1295 Leontopithecus rosalia 51.1 0.718 1295<	305
Hylobates lar 123.4 1.900 102 6.000 Lemur catta 45.1 2.678 25.6 2.678 Leontopitbecus rosalia 51.1 0.718 102 102 102 Leontopitbecus rosalia 51.1 0.718 102	263
Lemur catta 45.1 2.678 25.6 2.678 Leontopithecus rosalia 51.1 0.718 Lepilemur ruficaudatus 27.6 0.682 7.6 0.682 Loris tardigradus 14.8 0.284 6.6 0.322 Macaca fascicularis 400.9 7.100 74 5.500	181
Leontopitbecus rosalia 51.1 0.718 Lepilemur ruficaudatus 27.6 0.682 7.6 0.682 Loris tardigradus 14.8 0.284 6.6 0.322 Macaca fascicularis 400.9 7.100 74 5.500 Macaca fuscata 485.4 9.580 84 5.900 Macaca mulatta 231.9 5.380 110 8.000 Microcebus murinus 4.9 0.054 1.8 0.054 Nycticebus coucang 32.4 1.380 12.5 0.800 Otolemur crassicaudatus 47.6 0.950 10.3 0.850	166
Lepilemur ruficaudatus 27.6 0.682 7.6 0.682 Loris tardigradus 14.8 0.284 6.6 0.322 Macaca fascicularis 400.9 7.100 74 5.500 Macaca fuscata 485.4 9.580 84 5.900 Macaca mulatta 231.9 5.380 110 8.000 Microcebus murinus 4.9 0.054 1.8 0.054 Nycticebus coucang 32.4 1.380 12.5 0.800 Otolemur crassicaudatus 47.6 0.950 10.3 0.850	
Loris tardigradus 14.8 0.284 6.6 0.322 Macaca fascicularis 400.9 7.100 74 5.500 Macaca fascicularis 400.9 7.100 74 5.500 Macaca fuscata 485.4 9.580 84 5.900 Macaca mulatta 231.9 5.380 110 8.000 Microcebus murinus 4.9 0.054 1.8 0.054 Nycticebus coucang 32.4 1.380 12.5 0.800 Otolemur crassicaudatus 47.6 0.950 10.3 0.850	149
Macaca fascicularis 400.9 7.100 74 5.500 Macaca fascicularis 485.4 9.580 84 5.900 Macaca fuscata 485.4 9.580 84 5.900 Macaca mulatta 231.9 5.380 110 8.000 Microcebus murinus 4.9 0.054 1.8 0.054 Nycticebus coucang 32.4 1.380 12.5 0.800 Otolemur crassicaudatus 47.6 0.950 10.3 0.850	327.5
Macaca fuscata 485.4 9.580 84 5.900 Macaca mulatta 231.9 5.380 110 8.000 Microcebus murinus 4.9 0.054 1.8 0.054 Nycticebus coucang 32.4 1.380 12.5 0.800 Otolemur crassicaudatus 47.6 0.950 10.3 0.850	200
Macaca mulatta 231.9 5.380 110 8.000 Microcebus murinus 4.9 0.054 1.8 0.054 Nycticebus coucang 32.4 1.380 12.5 0.800 Otolemur crassicaudatus 47.6 0.950 10.3 0.850 Otolemur garnettii 47.8 1.028 275 10.3	223
Microcebus murinus 4.9 0.054 1.8 0.054 Nycticebus coucang 32.4 1.380 12.5 0.800 Otolemur crassicaudatus 47.6 0.950 10.3 0.850 Otolemur garnettii 47.8 1.028 275	159
Nycticebus coucang 32.4 1.380 12.5 0.800 Otolemur crassicaudatus 47.6 0.950 10.3 0.850 Otolemur garnettii 47.8 1.028 275 0.800	
Otolemur crassicaulatus 47.6 0.950 10.3 0.850 Otolemur garnettii 47.8 1.028 275 10.3	
Otolemur garnettii 47.8 1.028 275	195
Pan troglodytes 581.9 18.300 420 46.000	178
Papio anubis 342.9 9.500 205 26.000	207
Papio cynacephalus 668.9 14.300 195 19.000	184
Papio papio 297.3 6.230 190 18.000	
Papio ursinus 589.3 16.620 190 18.000	189.5
Perodicticus potto 41.3 1.000 14 1.150	190
Pongo pygmaeus 569.1 16.200 370 55.000	172.5
Propithecus verreauxi 86.8 3.080 26.7 3.480	200
Saguinus geoffroyi 50.5 0.500 10 3.800	263
Saimiri sciureus 68.8 0.850 22 6.800	323
<i>Tarsius syrichta</i> 8.9 0.113 350	
Varecia variegata 69.9 3.512 34.2 3.512	

 Table 1
 Comparative data on resting metabolic rate (RMR; kcal/day), body mass (kg), brain mass (g), and diet quality (DQ) in 41 primate species^a

^aData sources: References 8, 40, 42, 49, 58, 69, 70, 72, 79, and 85.

www.annualreviews.org • Brain Evolution and Human Metabolic Needs 313

Scaling (allometry): the change in size on biological measure with respect to another biological measure (often body size)

DQ: diet quality

mass (kg) for humans and the 40 other primate species from **Table 1**. It is clear that humans conform to the general primate scaling relationship between RMR and body weight, having RMRs that fall within 2% of the value predicted from the general primate relationship. The implication of this is that humans allocate a much larger share of our daily energy budget for brain metabolism than do other species.

The disproportionately high energy costs of our large brains are evident in Figure 2 (see color insert), which shows the scaling relationship between brain weight (grams) and RMR for humans and 35 other primate species (from Table 1) and 22 nonprimate mammalian species. The solid line denotes the best-fit regression for nonhuman primate species, and the dashed line denotes the best-fit regression for the nonprimate mammals. The slopes of the two regressions are similar (0.94 primates, 0.90 mammals; n.s.), whereas the y-intercepts are significantly different (-0.377 primates, -0.832 mammals; P < 0.01). Thus, at a given metabolic rate, primates have systematically larger brain sizes than those of other mammals, and humans, in turn, have larger brain sizes than do other primates. As a group, primates have brains that are approximately three times the size of brains of other mammals. Human brain sizes are some three times those of other primates.

The large allocation of our energy budget to brain metabolism raises the question of how humans are nutritionally able to accommodate the metabolic demands of our large brains. Recent work suggests that important dimensions of human nutritional biology are associated with the high energy demands of our large brains. It appears that humans consume diets that are denser in energy and nutrients in comparison with diets of other primates of similar size. Recent studies have shown that modern human foraging populations typically derive more than half of their dietary energy intake from animal foods, although considerable variation in diets exists (20, 41). In comparison, modern great apes obtain much of their diet from low-quality

plant foods. Gorillas derive more than 80% of their diet from fibrous foods such as leaves and bark (69). Even among common chimpanzees (*Pan troglodytes*), only about 5% to 10% of calories are derived from vertebrate animal foods (62, 78, 84). Field studies indicate that meat is a desirable and prized food item for many primate species. The low rates of consumption reflect the limited ability of chimpanzees and other primates to obtain large and consistent quantities of vertebrate foods because of high foraging costs (61). That is, the time and energy associated with pursuing game animals appear to be prohibitively high for most large-bodied primates.

Comparative dietary analyses of living primate species (including humans) are shown in **Figure 3** (see color insert), which plots dietary quality (DQ) as function of body mass (kg) for 33 different primate species (from **Table 1**). The DQ index was developed by Sailer et al. (72) and quantifies the energy and nutrient density of the diet based on the relative proportions of structural plant parts (s; e.g., leaves, stem, bark), reproductive plant parts (r; e.g., fruits, flowers), and animal foods (a; vertebrates and invertebrates):

$$DQ = s + 2r + 3.5a$$

The index ranges from a minimum of 100 (a diet of all leaves and/or structural plant parts) to 350 (a diet of all animal material).

Figure 3 shows that an inverse relationship exists between DQ and body mass (r = -0.59 total sample, -0.68 nonhuman primates only; P < 0.001). This tendency of larger primates to feed on lower-quality diets is something that is observed in other mammals (10, 39) and appears to be a consequence of the scaling relationship between energy requirements and body mass. As noted in **Figure 1**, the scaling coefficient between RMR and mass is less than one, implying that larger primates have proportionally lower metabolic rates than smaller ones. Large primates such as gorillas (*Gorilla gorilla*) and orangutans (*Pongo pygmaeus*) have high total energy requirements but relatively low massspecific needs (e.g., kcal/kg/day). They fulfill their energy needs by feeding on foods that are abundant but low in quality (e.g., leaves and foliage). Conversely, small animals [e.g., the pygmy marmoset (*Cebuella pygmaea*)] have low total energy requirements but very high mass-specific needs. They typically subsist on foods that are rich in calories and nutrients but relatively limited in abundance (e.g., saps, gums, and insects).

Humans, however, have substantially higher-quality diets than would be expected for a primate of our size. Note that the average diet for modern human foragers (based on dietary data from five modern human foraging populations; see 49) falls substantially above the regression line. Overall, the staple foods for *all* human societies are much more nutritionally dense than those of other largebodied primates. This higher-quality diet for humans relative to other large-bodied primates means that we need to eat a smaller volume of food to get the energy and nutrients we require.

Figure 4 (see color insert) shows relative brain size versus relative dietary quality for the 33 different primate species from Figure 3. Relative brain size for each species is measured as the standardized residual (z-score) from the primate brain versus body mass regression, and relative DQ is measured as the residual from the DQ versus body mass regression. There is a strong positive relationship (r =0.63; P < 0.001) between the amount of energy allocated to the brain and the caloric and nutrient density of the diet. Across all primates, larger brains require higher-quality diets. Humans fall at the positive extremes for both parameters, having the largest relative brain size (z = +3.27) and the highest quality diet (z = +2.05). Thus, the large, metabolically expensive human brain is partially offset by the consumption of an energy-dense and nutrient-rich diet. This relationship implies that the evolution of larger hominin brains would have necessitated the adoption of a sufficiently high-quality diet (including meat and energy-rich fruits) to support the increased metabolic demands of greater encephalization.

The relative size and morphology of the human GI tract also reflect our high-quality diet. Most large-bodied primates have expanded large intestines (colons), an adaptation to fibrous, low-quality diets (59). Humans, on the other hand, have small gut volumes for our size, with relatively enlarged small intestines and a smaller colon (3, 53, 75).

The enlarged colons of most large-bodied primates permits fermentation of low-quality plant fibers, allowing for extraction of additional energy in the form of volatile fatty acids (60, 63). In contrast, the GI morphology of humans (small colon and relatively enlarged small intestine) is more similar to a carnivore and reflects an adaptation to an easily digested, nutrient-rich diet (52, 53, 81).

Together, these comparative data suggest that the dramatic expansion of brain size over the course of human evolution likely would have required the consumption of a diet that was more concentrated in energy and nutrients than is typically the case for most large primates. This does not imply that dietary change was the driving force behind major brain expansion during human evolution. Rather, the available evidence indicates that a sufficiently high-quality diet was probably a necessary condition for supporting the metabolic demands associated with evolving larger hominin brains.

EVOLUTIONARY CHANGES IN BRAIN SIZE AND DIET

Trends in the Hominin Brain Size, Body Size, and Tooth Size

Over the past four million years, average brain size in the hominin lineage has more than tripled, increasing from approximately 400 cm³ in the earliest australopithecines to 1300–1400 cm³ in modern humans (57). However, the rates of evolutionary change in brain size have been highly variable over this Hominin: living humans and our fossil ancestors that lived after the last common ancestor between humans and apes

			Body weight		
Species	Geological age (mya)	Brain size (cm ³)	Male (kg)	Female (kg)	Postcanine tooth surface area (mm ²)
A. afarensis	3.9-3.0	438	45	29	460
A. africanus	3.0-2.4	452	41	30	516
A. boisei	2.3-1.4	521	49	34	756
A. robustus	1.9–1.4	530	40	32	588
Homo habilis (sensu strictu)	1.9–1.6	612	37	32	478
H. erectus (early)	1.8-1.5	863	66	54	377
H. erectus (late)	0.5-0.3	980	60	55	390
H. sapiens	0.4–0.0	1350	58	49	334

 Table 2 Geological ages (millions of years ago), brain size (cm³), estimated male and female body weights (kg), and postcanine tooth surface areas (mm²) for selected fossil hominid species

All data, except for *Homo erectus*, from (57). Early *H. erectus* brain size is the average of African specimens as presented in (56), Indonesian specimens from (5), and Georgian specimens from (31, 32). Data for late *H. erectus* are from (55).

period. Human evolution has been characterized by periods of slow increases in brain size alternating with periods of dramatic change. The human fossil record indicates that the first substantial burst of evolutionary change in hominin brain size occurred about 2.0 to 1.7 million years ago (mya) and was associated with the emergence and evolution of early members of our own genus, *Homo*.

Table 2 presents data on evolutionary changes in hominin brain size (cm³), estimated adult male and female body mass (kg), and posterior tooth area (mm²) (data from 5, 31, 32, 55–57). Hominin body masses were estimated from measurements of weightbearing joint surfaces using predictive equations derived from a diverse skeletal sample of modern humans (see 54). Posterior tooth areas are the summed surface areas of the premolar and molar teeth (57).

The australopithecines showed only modest brain size evolution from about 430 to 530 cm³ over more than two million years (from about 4 to 1.5 mya). With the evolution of the genus *Homo* there were substantial increases in encephalization, with brain sizes of over 600 cm³ in *Homo habilis* (at 1.9 to 1.6 mya) and 800 to 900 cm³ in early members of *Homo erectus* (at 1.8 to 1.5 mya). Although body sizes also increase with *H. erectus*, the changes in brain size are disproportionately greater than those in body mass. Thus, the level of encephalization we find with *H. erectus* is greater than that seen among any living nonhuman primate species today (49).

The changes in the craniofacial and dental anatomy of *H. erectus* suggest that these forms were consuming different foods from those consumed by its australopithecine relatives. During the evolution of the australopithecines, the total surface area of the grinding teeth increased dramatically from 460 mm² in *Australopithecus afarensis* to 756 mm² in *A. boisei*. In contrast, with the emergence of early *Homo* at approximately 2 mya, we see marked reductions in the posterior dentition. Postcanine tooth surface area is 478 mm² in *H. habilis* and 377 mm² in early *H. erectus*.

H. erectus also shows substantial reductions in craniofacial and mandibular robusticity relative to the australopithecines (91). Yet, despite having smaller teeth and jaws, *H. erectus* was a much bigger animal than the australopithecines, being humanlike in its stature, body mass, and body proportions (54, 55, 57, 71). Together, these features indicate that early *H. erectus* was consuming a richer, more calorically dense diet with less low-quality fibrous plant material. How the diet might have changed with the emergence of *H. erectus* is examined in the following section.

Australopithecus: genus of early hominins that existed in Africa between 4 and 1.2 mya

316 Leonard • Snodgrass • Robertson

Dietary Changes Associated with Brain Evolution in Early *Homo*

Increasing evidence suggests that the evolution of early Homo, particularly H. erectus, was associated with important changes in foraging patterns and dietary consumption compared with earlier hominin species. Recent analyses of dental microwear and stable carbon isotope levels of tooth enamel indicate that the australopithecines consumed a seasonally variable diet composed of both plant (e.g., fruits, seeds, grasses, and tubers) and some animal foods (76, 77, 83). Earlier interpretations of the so-called "robust" australopithecines (A. robustus and A. boisei)-with their massive jaws, robust faces, and large molar teeth-viewed these species as being dietary specialists that subsisted largely on seeds, nuts, and other hard objects common to the African savanna. However, the isotopic analyses now show that their diets were likely broader and more varied than previously thought (76, 77, 87). The consumption by australopithecines of limited quantities of animal foods (including invertebrates) is suggested by analogies with living primates (especially chimpanzees) and supported by stable isotope studies and association with putative bone tools likely used for termite extraction (7).

Paleontological and archaeological evidence indicates modest dietary change in earliest Homo (i.e., H. habilis); this species likely incorporated more animal foods in its diet, although the relative amounts obtained through hunting compared with scavenging is debated (12, 13, 36, 65). Evidence for dietary change in this species can be seen in the reduced masticatory functional complex (e.g., posterior tooth size); dental reduction in H. habilis reversed successive increases in cheek tooth size among the australopithecines (57). Technological advancements, such as the development of Oldowan industry tools, allowed easier processing of vertebrate carcasses and increased access to meat as well as energy- and nutrient-rich marrow and brains (74).

The evolution of *H. erectus* appears to be a major adaptive shift in human evolution. With the emergence of H. erectus in East Africa 1.8 mya we find (a) marked increases in brain and body size, (b) reductions of posterior tooth size and craniofacial robusticity, (c) the evolution of humanlike limb proportions, and (d) important changes in foraging/subsistence behavior (2, 4, 91, 92). These changes occurred within the context of large-scale climatic shifts (88, 95). The environment was becoming much drier, resulting in declines in forested areas and an expansion of open woodlands and grasslands (14, 26, 68, 95). Such a transformation of the African landscape would have strongly influenced the distribution of food resources for our hominin ancestors, making animal foods more abundant and thus an increasingly attractive food resource (9, 65). Using modern tropical ecosystems as our reference, we have found that although savanna/grasslands have much lower net primary (energetic) productivity than woodlands (4050 versus 7200 kcal/m²/yr), the level of herbivore productivity in savannas is almost three times that of the woodlands (10.2 versus 3.6 kcal/m²/yr) (50). Thus, fundamental changes in ecosystem structure 2.0 to 1.8 mya appear to have resulted in a net increase in the energetic abundance of grazing mammals (e.g., ungulates) on the E. African landscape. Such an increase would have offered an opportunity for hominins with sufficient behavioral and technological capability to exploit those resources.

The archeological record provides evidence that this occurred with *H. erectus*, as this species is associated with stone tools and the development of the first rudimentary hunting and gathering economy. Meat does appear to have been more common in the diet of *H. erectus* than it was in the australopithecines. H. erectus likely acquired mammalian carcasses through both hunting and confrontational scavenging (i.e., allowing other animal hunters to make the kill and then chasing them away from the carcass) (18, 65). In addition, the archaeological evidence **Oldowan:** the first stone tool technology in the human fossil record, characterized by simple flakes and choppers. First evident approximately 2.5 mya Acheulean: stone tool industry of the early and middle Pleistocene characterized by hand axes and cleavers. First evident 1.6 to 1.4 mya, associated with early *Homo*

DHA:

docosahexaenoic acid AA: arachidonic acid indicates that butchered animals were transported back to a central location (home base) where the resources were shared within foraging groups (18, 36, 66, 67). Increasingly sophisticated stone tools (i.e., the Acheulean industry) emerged approximately 1.6 to 1.4 mya, improving the ability of these hominins to process animal and plant materials (6). These changes in diet and foraging behavior would not have turned our hominin ancestors into carnivores; however, the addition of even modest amounts of meat to the diet (10% to 20% of dietary energy) combined with the sharing of resources that is typical of huntergatherer groups would have significantly increased the quality and stability of the diet of H. erectus.

Cordain and colleagues (22) have noted that beyond the energetic benefits, greater consumption of animal foods would have provided increased levels of key fatty acids that would have been necessary for supporting the rapid hominin brain evolution. Mammalian brain growth is dependent upon sufficient amounts of two long-chain polyunsaturated fatty acids: docosahexaenoic acid (DHA) and arachidonic acid (AA) (22, 24). Because the composition of all mammalian brain tissue is similar with respect to these two fatty acids, species with higher levels of encephalization have greater requirements for DHA and AA (24). It also appears that mammals have a limited capacity to synthesize these fatty acids from dietary precursors. Consequently, dietary sources of DHA and AA were likely limiting nutrients that constrained the evolution

of larger brain size in many mammalian lineages (23, 24).

Cordain and colleagues (22) have demonstrated that wild plant foods available on the African savanna (e.g., tubers, nuts) contain, at most, trace amounts of AA and DHA, whereas muscle tissue and organ meat of wild African ruminants provide moderate to high levels of these key fatty acids. As shown in Table 3, brain tissue is a rich source of both AA and DHA, whereas liver and muscle tissues are good sources of AA and moderate sources of DHA. Other good sources of AA and DHA are freshwater fish and shellfish (17, 22, 24). Cunnane and colleagues (17, 25) have suggested that the major increases in hominin encephalization were associated with systematic use of aquatic (marine, riverine, or lacustrian) resources. However, there is little archeological evidence for the systematic use of aquatic resources until much later in human evolution (45).

An alternative strategy for increasing dietary quality in early *Homo* has been proposed by Wrangham and colleagues (93, 94). These authors argue that the controlled use of fire for cooking allowed early *Homo* to improve the nutritional density of their diet. They note that the cooking of savanna tubers and other plant foods would have served to both soften them and increase their energy/nutrient bioavailability. In their raw form, the starch in roots and tubers is not absorbed in the small intestine and is passed through the body as nondigestible carbohydrate (30, 82). However, when heated, the

Table 3 Energy (kcal), fat (g), protein (g), arachidonic acid (AA), and docosahexaenoic acid (DHA) contents of African ruminant, fish, and wild plant foods per 100 grams. Data derived from (22)

Food item	Energy (kcal)	Fat (g)	Protein (g)	AA (mg)	DHA (mg)
African ruminant (brain)	126	9.3	9.8	533	861
African ruminant (liver)	159	7.1	22.6	192	41
African ruminant (muscle)	113	2.1	22.7	152	10
African ruminant (fat)	745	82.3	1.0	20-180	trace
African fish	119	4.5	18.8	270	549
Wild tuber/roots	96	0.5	2.0	0	0
Mixed wild plants	129	2.8	4.1	0	0

318 Leonard • Snodgrass • Robertson

starch granules swell and are disrupted from the cell walls. This process, known as gelatinization, makes the starch much more accessible to breakdown by digestive enzymes (34). Thus, cooking increases the nutritional quality of tubers by making more of the carbohydrate energy available for biological processes.

Although cooking, which served to increase dietary digestibility and quality, is clearly an important innovation in hominin evolution, there is very limited evidence for the controlled use of fire by hominins before 1.5 mya (11, 15, 64). The more widely held view is that the use of fire and cooking did not occur until later in human evolution, at 200,000 to 250,000 years ago (80, 90). Moreover, nutritional analyses of wild tubers used by modern foraging populations (e.g., 16, 73, 89) suggest that the energy content of these resources is markedly lower than that of animal foods, even after cooking (22). Unlike animal foods, tubers are also devoid of both DHA and AA (22; see Table 3). Consequently, major questions remain about whether cooking and the heavy reliance on roots and tubers were important forces for promoting rapid brain evolution with the emergence of early Homo.

Overall, the available evidence seems to best support a mixed dietary strategy in early Homo that involved the consumption of larger amounts of animal foods in comparison with the australopithecines. Ungar and colleagues (87) recently suggested that early Homo likely pursued a flexible and versatile subsistence strategy that would have allowed them to adapt to the patchy and seasonally variable distribution of food resources on the African savanna. They note that such a model is more plausible than are ones proposing heavy reliance on one particular type of resource (e.g., meat or tubers). This is indeed true; however, what appears to be happening with early Homo-especially with H. erectus-is the development of a more stable and effective way of extracting resources from the environment. The increase in dietary quality and stability

was likely achieved partly through changes in diet composition (22, 49) and partly through social and behavioral changes like food sharing and perhaps division of foraging tasks (36, 38, 41). This greater nutritional stability provided a critical foundation for fueling the energy demands of larger brain sizes.

BRAIN EVOLUTION AND HUMAN BODY COMPOSITION

In addition to improvements in dietary quality, the increased metabolic cost of larger brain size in human evolution also appears to have been supported by changes in body composition. Humans allocate a substantially larger share of their daily energy budget to their brains than do other primates or other mammals, which implies that the size and metabolic demands of certain other organs/organ systems may be relatively reduced in humans compared with other species. Thus, the critical question is, which organs have been reduced or altered in their relative size over the course of human evolution to compensate for the expansion of brain size?

Analyses of human and primate body composition offer possible answers to this question. Aiello (1) and Aiello & Wheeler (3) have argued that the increased energy demands of the human brain were accommodated by the reduction in size of the GI tract. Since the intestines are similar to the brain in having very high energy demands (so-called expensive tissues), the reduction in size of the large intestines of humans relative to other primates is thought to provide the necessary energy "savings" required to support elevated brain metabolism. Aiello & Wheeler (3) have shown that among a sample of 18 primate species (including humans), increased brain size was associated with reduced gut size. However, recent analyses by Snodgrass et al. (75) have failed to demonstrate significant differences in GI size between primates and nonprimate mammals that are predicted from the expensive tissue hypothesis. Thus, while it is clear that humans have relatively small GI sizes for their body mass, questions remain about the extent to which reductions in GI size helped to balance the increased metabolic costs associated with expansion of brain size during the course of human evolution. The reduced GI size in humans instead may be the direct consequence of improvements in DQ over the course of human evolution.

Leonard and colleagues (51) and Kuzawa (46) have suggested that differences in muscle and fat mass between humans and other primates may also account for variation in the budgeting of metabolic energy. Relative to other primates and other mammals, humans have lower levels of muscle mass and higher levels of body fatness (46, 51, 65). The relatively high levels of adiposity in humans have two important metabolic implications for brain metabolism. First, because fat has lower energy requirements than that of muscle tissue, replacing muscle mass with fat mass results in energy savings that can be allocated to the brain. Additionally, fat provides a ready source of stored energy that can be drawn upon during periods of limited food availability. Consequently, the higher levels of body fat in humans may also help to support larger brain size by providing stored energy to buffer against environmental fluctuations in nutritional resources.

The importance of body fat is particularly notable in human infants, which have both high brain-to-body weight ratios and high levels of body fatness (46). **Table 4** shows agerelated changes in body weight (kg), brain weight (g), fatness (%), RMR (kcal/day), and percent of RMR allocated to the brain for humans from birth to adulthood. We see that in infants, brain metabolism accounts for upward of 60% of RMR. Human infants are also considerably fatter than infants of other mammalian species (46). Body fatness in human infants is approximately 15% to 16% at birth, and continues to increase to 25% to 26% during the first 12 to 18 months of postnatal growth. Fatness then declines to about 15% by early childhood (27). Thus, during early human growth and development, it appears that body fatness is highest during the periods of the greatest metabolic demand of the brain.

It is likely that fundamental changes in body composition (i.e., the relative sizes of different organ systems) during the course of hominin evolution allowed for the expansion of brain size without substantial increases in the total energy demands for the body. At present, we do not know which alterations were the most critical for accommodating brain expansion. Variation in body composition both within and between primate species is still not well understood. Our knowledge of variation in body composition among humans is based largely on data from populations of the industrialized world. Consequently, more and better data on interspecific and ontogenetic variation in primate and human body composition are necessary to further resolve these issues

Age	Body weight (kg)	Brain weight (g)	Body fat (%)	RMR (kcal/day)	BrMet (%)
Newborn	3.5	475	16	161	87
3 months	5.5	650	22	300	64
18 months	11.0	1045	25	590	53
5 years	19.0	1235	15	830	44
10 years	31.0	1350	15	1160	34
Adult male	70.0	1400	11	1800	23
Adult female	50.0	1360	20	1480	27

Table 4 Body weight (kg), brain weight (g), percent body fat (%), resting metabolic rate (RMR; kcal/day), and percent of RMR allocated to brain metabolism (BrMet, %) for humans from birth to adulthood^a

^aAll data are from (37), except for percent body fat data for children 18 months and younger, which are from (27).

320 Leonard • Snodgrass • Robertson

New imaging techniques such as magnetic resonance imaging and positron emission tomography scans offer the potential to directly explore variation in organ weight and organ-specific energy demands in living humans and primates. For example, Gallagher et al. (33) recently used magnetic resonance imaging technology to measure how differences in organ weights contribute to ethnic differences in RMRs among living humans. These authors demonstrated that the significant differences in RMR between their African American and Euro-American samples could be accounted for by differences in the summed weight of the most metabolically expensive organs (liver, heart, spleen, kidneys, and brain). Similarly, Chugani (19) utilized positron emission tomography scans to quantify changes in glucose utilization in the human brain from birth to adulthood. His findings suggest that the extremely high metabolic costs of brain metabolism characteristic of early human life (as outlined in Table 4) may extend further into childhood than previously realized. Together, these studies highlight the potential use of new imaging techniques for better understanding how interspecific variation in body composition contributes to differences in metabolic rate.

CONCLUSIONS

The evolution of large human brain size has had important implications for the nutritional biology of our species. Our large brains are energetically expensive, yet, paradoxically, our overall metabolic requirements are similar to those of any comparably sized mammal. As a consequence, humans expend a relatively larger proportion of their resting energy budget on brain metabolism than do other primates or nonprimate mammals.

Comparative analyses of primate dietary patterns indicate that the high costs of large human brains are supported, in part, by diets that are relatively rich in energy and other nutrients. Among living primates, the relative proportion of metabolic energy allocated to the brain is positively correlated with dietary quality. Humans fall at the positive end of this relationship, having both a very high-quality diet and a large brain.

Greater encephalization also appears to have consequences for other aspects of body composition, most notably the GI mass, muscularity, and adiposity. Relative to other primates, human have smaller GI tracts and a relatively reduced colon. This type of gut is consistent with adaptation to a diet that is relatively high in energy and nutrients and is easy to digest.

In addition, humans appear to be relatively undermuscled (i.e., less skeletal muscle) and over fat compared with other primates of similar size. The relatively high levels of adiposity in humans are particularly notable in infancy. These greater levels of body fatness and reduced levels of muscle mass allow human infants to accommodate the growth of their large brains in two important ways: (*a*) by having a ready supply of stored energy to feed the brain and (*b*) by reducing the total energy costs of the rest of the body.

The human fossil record indicates that major changes in both brain size and diet occurred in association with the emergence of early members of the genus Homo between 2.0 and 1.7 mya in Africa. With the evolution of early H. erectus 1.8 mya, we find evidence of an important adaptive shift-the evolution of the first hunting and gathering economy, characterized by greater consumption of animal foods, transport of food resources to home bases, and sharing of food within social groups. H. erectus was humanlike in body size and proportions and had a brain size beyond that seen in nonhuman primates, approaching the range of modern humans. In addition, the reduced size of the face and grinding teeth of H. erectus, coupled with its more sophisticated tool technology, suggest that these hominins were consuming a higher quality and more stable diet that would have helped to fuel the increases in brain size. Consequently, although dietary change was not the prime force responsible for the evolution of large human brain size, improvements in dietary quality appear to have been a necessary condition for promoting encephalization in the human lineage. took place with emergence of *Homo*. In addition, the application of new biomedical imaging techniques offers the potential to directly explore how intra- and interspecific variation in body composition may contribute to variation in metabolic rates.

Further research is needed to better understand the nature of the dietary changes that

SUMMARY POINTS

- 1. Our large brains are energetically expensive, yet paradoxically our overall metabolic requirements are similar to those of any comparably sized mammal. Consequently, humans expend a relatively larger proportion of their resting energy budget on brain metabolism than do other primates or nonprimate mammals.
- 2. Comparative analyses of living primate species show that the relative proportion of metabolic energy allocated to the brain is positively correlated with dietary quality. Humans fall at the positive end of this relationship, having both a very high-quality diet and a large brain. This suggests that large human brains are supported, in part, by diets that are relatively rich in energy and other nutrients.
- 3. Compared with other primates, humans have smaller overall gastrointestinal tracts with a relatively reduced colon. This type of gut is consistent with adaptation to a diet that is relatively high in energy and nutrients and is easy to digest.
- 4. Humans have relatively lower levels of muscularity and higher levels of adiposity than do other primates of similar size. High levels of adiposity in humans are particularly notable in infancy. Greater body fatness and lower muscle mass allow human infants to accommodate the growth of their large brains by having a ready supply of stored energy, reducing the total energy costs of the rest of the body.
- 5. The human fossil record indicates that major changes in both brain size and diet occurred in association with the emergence of early members of the genus *Homo* between 2.0 and 1.7 mya in Africa. With the evolution of early *H. erectus* 1.8 mya, we find evidence of an important adaptive shift—the evolution of the first hunting and gathering economy, characterized by greater consumption of animal foods, transport of food resources to home bases, and sharing of food within social groups. Improvements in diet quality with *H. erectus* appear to have been important for fueling rapids rates of encephalization.
- 6. Consumption of more animal foods with early *Homo* was likely important for providing high levels of key long-chain polyunsaturated fatty acids (docosahexaenoic acid and arachidonic acid) that are necessary for brain growth.

ACKNOWLEDGMENTS

We are grateful to S.C. Antón and C.W. Kuzawa for discussions about this research.

LITERATURE CITED

1. Aiello LC. 1997. Brains and guts in human evolution: the expensive tissue hypothesis. *Braz. 7. Genet.* 20:141–48

322 Leonard • Snodgrass • Robertson

1. Posits that the high metabolic costs of large human brains have been partially offset by the reductions in the size of our gastrointestinal tracts.

- Aiello LC, Key C. 2002. Energetic consequences of being a *Homo erectus* female. Am. J. Hum. Biol. 14:551–65
- 3. Aiello LC, Wheeler P. 1995. The expensive-tissue hypothesis: the brain and the digestive system in human and primate evolution. *Curr. Anthropol.* 36:199–221
- 4. Antón SC. 2003. A natural history of Homo erectus. Yrbk. Phys. Anthropol. 46:126-70
- Antón SC, Swisher CC III. 2001. Evolution of cranial capacity in Asian Homo erectus. In A Scientific Life: Papers in Honor of Dr. T. Jacob, ed. E Indriati, pp. 25–39. Yogyakarta, Indonesia: Bigraf
- Asfaw B, Beyene Y, Suwa G, Walter RC, White TD, et al. 1992. The earliest Acheulean from Konso-Gardula. *Nature* 360:732–35
- Backwell LR, d'Errico F. 2001. Evidence of termite foraging by Swartkrans early hominids. Proc. Nat. Acad. Sci. USA 98:1358–63
- Bauchot R, Stefan H. 1969. Encephalisation et niveau evolutif ches les simiens. Mammalia 33:225–75
- Behrensmeyer K, Todd NE, Potts R, McBrinn GE. 1997. Late Pliocene faunal turnover in the Turkana basin, Kenya and Ethiopia. *Science* 278:1589–94
- 10. Bell RH. 1971. A grazing ecosystem in the Serengeti. Sci. Am. 225(1):86-93
- Bellomo RV. 1994. Methods of determining early hominid behavioral activities associated with the controlled use of fire at FxJj 20 Main, Koobi Fora. *J. Hum. Evol.* 27:173–95
- Blumenschine RJ. 1987. Characteristics of the early hominid scavenging niche. Curr. Anthropol. 28:383–407
- Blumenschine RJ, Cavallo JA, Capaldo SD. 1994. Competition for carcasses and early hominid behavioral ecology: a case study and conceptual framework. *J. Hum. Evol.* 27:197– 213
- Bobe R, Behrensmeyer AK. 2002. Faunal change, environmental variability and late Pliocene hominin evolution. *7. Hum. Evol.* 42:475–97
- Brain CK, Sillen A. 1988. Evidence from the Swartkrans cave for the ealiest use of fire. Nature 336:464–66
- Brand-Miller JC, Holt SHA. 1998. Australian aboriginal plant foods: a consideration of their nutritional composition and health implications. *Nutr. Res. Rev.* 11:5–23
- Broadhurst CL, Cunnane SC, Crawford MA. 1998. Rift Valley lake fish and shellfish provided brain-specific nutrition for early Homo. Br. J. Nutr. 79:3–21
- Bunn HT. 2006. Meat made us human. In Evolution of the Human Diet: The Known, the Unknown, and the Unknowable, ed. PS Unger, pp. 191–211. New York: Oxford Univ. Press
- Chugani HT. 1998. A critical period of brain development: studies of cerebral glucose utilization with PET. *Prevent. Med.* 27:184–88
- Cordain L, Brand-Miller J, Eaton SB, Mann N, Holt SHA, Speth JD. 2000. Plant to animal subsistence ratios and macronutrient energy estimations in world-wide hunter-gatherer diets. Am. J. Clin. Nutr. 71:682–92
- Cordain L, Eaton SB, Sebastian A, Mann N, Lindberg S, et al. 2005. Origins and evolution of the Western diet: health implications for the 21st century. *Am. J. Clin. Nutr.* 81:341–54
- Cordain L, Watkins BA, Mann NJ. 2001. Fatty acid composition and energy density of foods available to African hominids. *World Rev. Nutr. Diet.* 90:144–61
- Crawford MA. 1992. The role of dietary fatty acids in biology: their place in the evolution of the human brain. *Nutr. Rev.* 50:3–11
- Crawford MA, Bloom M, Broadhurst CL, Schmidt WF, Cunnane SC, et al. 1999. Evidence for unique function of docosahexaenoic acid during the evolution of the modern human brain. *Lipids* 34:S39–47

- 25. Cunnane SC, Crawford MA. 2003. Survival of the fattest: fat babies were the key to evolution of the large human brain. *Comp. Biochem. Physiol. A* 136:17–26
- deMenocal PB. 2004. African climate change and faunal evolution during the Pliocene-Pleistocene. *Earth Planet. Sci. Lett.* 220:3–24
- Dewey KG, Heinig MJ, Nommsen LA, Peerson JM, Lonnerdal B. 1993. Breast-fed infants are leaner than formula-fed infants at 1 year of age: the Darling Study. *Am. J. Clin. Nutr.* 52:140–45
- Eaton SB. 2006. The ancestral human diet: What was it and should it be a paradigm for contemporary nutrition? *Proc. Nutr. Soc.* 65:1–6
- 29. Eaton SB, Konner MJ. 1985. Paleolithic nutrition: a consideration of its nature and current implications. *New Engl. J. Med.* 312:283–89
- 30. Englyst KN, Englyst HN. 2005. Carbohydrate bioavailability. Br. 7. Nutr. 94:1-11
- Gabunia L, Vekua A, Lordkipanidze D, Swisher CC, Ferring R, et al. 2000. Earliest Pleistocene cranial remains from Dmanisi, Republic of Georgia: taxonomy, geological setting, and age. *Science* 288:1019–25
- Gabunia L, Antón SC, Lordkipanidze D, Vekua A, Justus A, Swisher CC III. 2001. Dmanisi and dispersal. *Evol. Anthropol.* 10:158–70
- Gallagher D, Albu J, He Q, Heshka S, Boxt L, et al. 2006. Small organs with a high metabolic rate explain lower resting energy expenditure in African American than in white adults. *Am. 7. Clin. Nutr.* 83:1062–67
- García-Alonso A, Goñi I. 2000. Effect of processing on potato starch: in vitro availability and glycemic index. *Nahrung* 44:19–22
- 35. Garn SM, Leonard WR. 1989. What did our ancestors eat? Nutr. Rev. 47:337-45
- 36. Harris JWK, Capaldo S. 1993. The earliest stone tools: their implications for an understanding of the activities and behavior of late Pliocene hominids. In *The Use of Tools by Human and Nonhuman Primates*, ed. A Berthelet, J Chavaillon, pp. 196–220. Oxford: Oxford Sci.
- Holliday MA. 1986. Body composition and energy needs during growth. In *Human Growth: A Comprehensive Treatise*, *Volume 2*, ed. F. Falkner, JM Tanner, pp. 101–17, New York: Plenum. 2nd ed.
- Isaac GL. 1978. Food sharing and human evolution: archaeological evidence from the Plio-Pleistocene of East Africa. *J. Anthropol. Res.* 34:311–25
- Jarman PJ. 1974. The social organization of antelope in relation to their ecology. *Behaviour* 58:215–67
- 40. Jerison HJ. 1973. The Evolution of the Brain and Intelligence. New York: Academic
- Kaplan H, Hill K, Lancaster J, Hurtado AM. 2000. A theory of life history evolution: diet, intelligence and longevity. *Evol. Anthropol.* 9:156–85
- 42. Kappeler PM. 1996. Causes and consequences of life-history variation among strepsirhine primates. *Am. Nat.* 148:868–91
- 43. Kety SS. 1957. The general metabolism of the brain in vivo. In *Metabolism of the Central Nervous System*, ed. D Richter, pp. 221–37. New York: Pergammon
- 44. Kleiber M. 1961. The Fire of Life. New York: Wiley
- Klein RG. 1999. The Human Career: Human Biological and Cultural Origins. Chicago: Univ. Chicago Press. 2nd ed.
- Kuzawa CW. 1998. Adipose tissue in human infancy and childhood: an evolutionary perspective. *Yrbk. Phys. Anthropol.* 41:177–209
- 47. Leonard WR. 2002. Food for thought: dietary change was a driving force in human evolution. *Sci. Am.* 287(6):106–15

324 Leonard • Snodgrass • Robertson

29. One of the first papers to consider human nutrition from an evolutionary perspective. Argues that many of today's common diseases stem from diets that differ from those of our prehistoric ancestors.

46. Demonstrates the adaptive value of body fat in human infants for buffering against disease and supporting the high energetic costs of large, growing brains.

47. Uses an energetics approach for understanding major trends in human evolution, such as bipedality, encephalization, and the expansion of hominins from Africa.

- Leonard WR, Robertson ML. 1992. Nutritional requirements and human evolution: a bioenergetics model. Am. J. Hum. Biol. 4:179–95
- Leonard WR, Robertson ML. 1994. Evolutionary perspectives on human nutrition: the influence of brain and body size on diet and metabolism. *Am. J. Hum. Biol.* 6:77–88
- Leonard WR, Robertson ML. 1997. Comparative primate energetics and hominid evolution. Am. J. Phys. Anthropol. 102:265–81
- Leonard WR, Robertson ML, Snodgrass JJ, Kuzawa CW. 2003. Metabolic correlates of hominid brain evolution. Comp. Biochem. Physiol. A 135:5–15
- 52. Martin RD. 1989. Primate Origins and Evolution: A Phylogenetic Reconstruction. Princeton, NJ: Princeton Univ. Press
- Martin RD, Chivers DJ, MacLarnon AM, Hladik CM. 1985. Gastrointestinal allometry in primates and other mammals. In *Size and Scaling in Primate Biology*, ed. WL Jungers, pp. 61–89. New York: Plenum
- McHenry HM. 1992. Body size and proportions in early hominids. Am. J. Phys. Anthropol. 87:407–31
- McHenry HM. 1994a. Tempo and mode in human evolution. Proc. Natl. Acad. Sci. USA 91:6780–86
- McHenry HM. 1994b. Behavioral ecological implications of early hominid body size. *J. Hum. Evol.* 27:77–87
- McHenry HM, Coffing K. 2000. Australopithecus to Homo: transformations in body and mind. *Annu. Rev. Anthropol.* 29:125–46
- McNab BK, Wright PC. 1987. Temperature regulation and oxygen consumption in the Philippine tarsier *Tarsius syrichta*. *Physiol. Zoo.* 60:596–600
- Milton K. 1987. Primate diets and gut morphology: implications for hominid evolution. In *Food and Evolution: Toward a Theory of Human Food Habits*, ed. M Harris, EB Ross, pp. 93–115. Philadelphia, PA: Temple Univ. Press

60. Milton K. 1993. Diet and primate evolution. Sci. Am. 269(2):86-93

- Milton K. 1999. A hypothesis to explain the role of meat-eating in human evolution. *Evol.* Anthropol. 8:11–21
- Milton K. 2003. The critical role played by animal source foods in human (*Homo*) evolution. *7. Nutr*. 133:3886–92S
- Milton K, Demment MW. 1988. Digestion and passage kinetics of chimpanzees fed highand low-fiber diets and comparison with human data. *J. Nutr.* 118:1082–88
- 64. Pennisi E. 1999. Did cooked tubers spur the evolution of big brains? Science 283:2004-5
- Plummer T. 2004. Flaked stones and old bones: biological and cultural evolution at the dawn of technology. *Yrbk. Phys. Anthropol.* 47:118–64
- 66. Potts R. 1988. Early Hominid Activities at Olduvai. New York: Aldine
- Potts R. 1998. Environmental hypotheses of hominin evolution. Yrbk. Phys. Anthropol. 41:93–136
- Reed K. 1997. Early hominid evolution and ecological change through the African Plio-Pleistocene. *J. Hum. Evol.* 32:289–322
- 69. Richard AF. 1985. Primates in Nature. New York: Freeman
- 70. Rowe N. 1996. The Pictorial Guide to Living Primates. New York: Pogonias
- Ruff CB, Trinkaus E, Holliday TW. 1997. Body mass and encephalization in Pleistocene Homo. *Nature* 387:173–76
- Sailer LD, Gaulin SJC, Boster JS, Kurland JA. 1985. Measuring the relationship between dietary quality and body size in primates. *Primates* 26:14–27

52. Synthesizes research on primate and human evolution, drawing on a rich body of comparative data on primate biology.

60. Examines the evolution of primate and human diets, drawing on comparative studies of primate ecology.

- Schoeninger MJ, Bunn HT, Murray SS, Marlett JA. 2001. Nutritional composition of some wild plant foods and honey used by Hadza foragers of Tanzania. *J. Food Comp. Anal.* 14:3–13
- 74. Semaw S, Rogers MJ, Quade J, Renne PR, Butler RF, et al. 2003. 2.6-million-year-old stone tools and associated bones from OGS-6 and OGS-7, Gona, Afar, Ethiopia. *J. Hum. Evol.* 45:169–77
- 75. Snodgrass JJ, Leonard WR, Robertson ML. 2007. Energetics of encephalization in early hominids. In *The Evolution of Hominid Diets: Integrating Approaches to the Study of Palaeolithic Subsistence*, ed. M Richards, JJ Hublin. New York: Springer. In press
- Sponheimer M, Lee-Thorp J, de Ruiter DJ, Codron D, Codron J, et al. 2005. Hominins, sedges, and termites: new carbon isotope data from Sterkfontein valley and Kruger National Park. *J. Hum. Evol.* 48:301–12
- Sponheimer M, Passey BH, de Ruiter DJ, Guatelli-Steinberg D, Cerling TE, Lee-Thorp JA. 2006. Isotopic evidence for dietary variability in the early hominin *Paranthropus robustus*. *Science* 314:980–82
- Stanford CB. 1996. The hunting ecology of wild chimpanzees: implications for the evolutionary ecology of Pliocene hominids. Am. Anthropol. 98:96–113
- 79. Stephan H, Frahm H, Baron G. 1981. New and revised data on volumes of brain structures in insectivores and primates. *Folia Primatol*. 35:1–29
- 80. Straus LG. 1989. On early hominid use of fire. Curr. Anthropol. 30:488-91
- Sussman RW. 1987. Species-specific dietary patterns in primates and human dietary adaptations. In *Evolution of Human Behavior: Primate Models*, ed. W Kinzey, pp. 151–79. Albany, NY: SUNY Press
- Tagliabue A, Raben A, Heijnen ML, Duerenberg P, Pasquali E, Astrup A. 1995. The effect of raw potato starch on energy expenditure and substrate oxidation. *Am. J. Clin. Nutr.* 61:1070–75
- Teaford MF, Ungar PS. 2000. Diet and the evolution of the earliest human ancestors. *Proc. Natl. Acad. Sci. USA* 97:13506–11
- 84. Teleki G. 1981. The omnivorous diet and eclectic feeding habits of the chimpanzees of Gombe National Park. In *Omnivorous Primates*, ed. RSO Harding, G Teleki, pp. 303–43. New York: Columbia Univ. Press
- Thompson SD, Power ML, Rutledge CE, Kleiman DG. 1994. Energy metabolism and thermoregulation in the golden lion tamarin (*Leontopithecus rosalia*). *Folia Primatol*. 63:131– 43
- 86. Ungar PS, ed. 2007. Evolution of the Human Diet: The Known, the Unknown, and the Unknowable. New York: Oxford Univ. Press
- 87. Ungar PS, Grine FE, Teaford MF. 2006. Diet in early *Homo*: a review of the evidence and a new model of adaptive versatility. *Annu. Rev. Anthropol.* 35:209–28
- Vrba ES. 1995. The fossil record of African antelopes relative to human evolution. In Paleoclimate and Evolution, With Emphasis on Human Origins, ed. ES Vrba, GH Denton, TC Partridge, LH Burkle, pp. 385–424. New Haven, CT: Yale Univ. Press
- Wehmeyer AS, Lee RB, Whiting M. 1969. The nutrient composition and dietary importance of some vegetable foods eaten by the !Kung bushmen. S. Afr. Med. J. 95:1529–30
- Weiner S, Qunqu X, Goldberg P, Liu J, Bar-Yosef O. 1998. Evidence for the use of fire at Zhoukoudian, China. *Science* 281:251–53
- 91. Wolpoff MH. 1999. Paleoanthropology. Boston: McGraw-Hill. 2nd ed.
- 92. Wood B, Collard M. 1999. The human genus. Science 284:65-71

326 Leonard • Snodgrass • Robertson

86. Provides an authoritative overview of research on human nutritional evolution from the fields of paleontology, archaeology, primatology and comparative human biology.

- Wrangham RW, Conklin-Brittain NL. 2003. Cooking as a biological trait. Comp. Biochem. Physiol. A Mol. Integr. Physiol. 136:35–46
- 94. Wrangham RW, Jones JH, Laden G, Pilbeam D, Conklin-Brittain NL. 1999. The raw and the stolen: cooking and the ecology of human origins. *Curr. Anthropol.* 40:567–94
- Wynn JG. 2004. Influence of Plio-Pleistocene aridification on human evolution: evidence from paleosols from the Turkana Basin, Kenya. Am. J. Phys. Anthropol. 123:106–18



Log-Log plot of resting metabolic rate (RMR; kcal/day) versus body mass (kg) for 41 species of primates (including humans). Humans conform to the general primate scaling relationship $[RMR = 55(Wt^{0.81})]$. Adapted from Reference 51.



Log-Log plot of brain weight (BW; g) versus resting metabolic rate (RMR) (kcal/day) for humans, 35 other primate species, and 22 species of nonprimate mammals. The primate regression line (*solid*) is elevated systematically and significantly above the nonprimate mammal regression (*dasbed*) (y-intercepts = -0.377 primates, -0.832 mammals; P < 0.01). The scaling relationships for nonprimate mammals are BW = 0.14 (RMR^{0.90}); primates, BW = 0.42 (RMR^{0.94}). Thus, for a given RMR, primates have brain sizes that are approximately three times those of other mammals, and humans have brains that are three times those of other primates.



Plot of diet quality (DQ) versus log-body mass for 33 primate species. DQ is inversely related to body mass (r = -0.59 total sample, -0.68 nonhuman primates only; P < 0.001), indicating that smaller primates consume relatively higher-quality diets. Humans have systematically higher-quality diets than predicted for their size.



Plot of relative brain size versus relative diet quality for 33 primate species. Primates with higher-quality diets for their size have relatively larger brain size (r = 0.63; P < 0.001). Humans represent the positive extremes for both measures, having large brain:body size and a substantially higher-quality diet than would be expected for their size.